

RECENT TRENDS IN THE UTILIZATION OF MEDICINAL PLANTS AS GROWTH PROMOTERS IN POULTRY NUTRITION – A REVIEW

R.A. Oluwafemi¹, A. Isiaka Olawale¹, J.O. Alagbe^{2*}

¹Department of Animal Science, Faculty of Agriculture, University of Abuja, Nigeria

²Department of Animal Nutrition, Sumitra Research Farm, Gujarat, India

Abstract. Plants are one of the numerous gifts of nature, they are loaded with several bioactive chemicals or phytochemicals as part of their normal metabolic activities. The beneficial effects of phytochemicals could be attributed to their antimicrobial, antiviral, antifungal and antioxidant properties. The use of medicinal plants (herbs, spices and their extracts) as organic alternatives have increased due to the increasing awareness on food safety and the dangers on the indiscriminate use of antibiotics. Herbs are cheap, easily available and produce no toxic residue on final products from animals when compared with antibiotics. Phytochemicals (such as flavonoids, phenols, saponin, alkaloids etc.) performs multiple biological activities in animals, such activities include improving performance, increased proliferations of immune cells, relief from intestinal challenge, reduction in oxidative stress, reduced mortality and increased antibody titers. The efficacy of these bioactive chemicals and results obtained from animals depends on extraction method, stage and age of plants, geographical location, species or breeds of animals, management methods and concentrations administered. So many potential abound in the use of medicinal plants and it was even recommended by the European Union (2006) as botanical alternative to antibiotics. Therefore, this review focuses on ways to bridge the gap between food safety and animal production.

Keywords: medicinal plants, phytochemical, bioactive chemicals, antibiotics.

***Corresponding Author:** J.O. Alagbe, Department of Animal Nutrition, Sumitra Research Farm, Gujarat, India, e-mail: demsonfarms@yahoo.com

Received: 13 August 2019;

Accepted: 12 March 2020;

Published: 15 April 2020.

1. Introduction

The use of medicinal plants in livestock production is gaining more recognition as a result of numerous efforts in phasing out antibiotics due to public reservations regarding the development of antibiotics – resistant bacteria and residual effects in food (Ekunseitan *et al.*, 2016). These development has prompted the European Union in 2006 to place a ban on the use of antibiotics, medicinal plants are however used as botanical or alternative to antibiotics in animal husbandry. The worldwide interest on herbal products has grown significantly as described by Verigi *et al.* (2003), cattle, goat, sheep, horses and pigs represents about 70% of the animals treated with herbal remedies followed by poultry (9.1%), dogs (5.3%) and rabbits (4.3%). Medicinal plants are mostly used especially by rural and small scale farmers because they are cheap, accessible and effective.

Recent researchers have suggested the use of probiotics, prebiotics, organic acids and plant extracts and essential oils as natural alternatives for replacing antibiotic growth promoters (AGPs) in poultry feed because they have antimicrobial, antiviral, anti-parasitic, antifungal, antioxidant and anti-helminthic properties due to the presence of phytochemical or bioactive chemicals in plants (Fulton *et al.*, 2002; Gopal, 2014). To

gain the advantageous effect of medicinal plants, they can be added to feed as dried plants or part of plants as extracts (Tamara *et al.*, 2009). The content of medicinal plants and their final products may vary widely depending on the parts used (seeds, leaves etc.), extraction method, stage and age of maturity, geographical origin, storage conditions and harvesting seasons (Burt, 2004). Medicinal plants have been shown to offer wide range of activities, including animal performance and nutrient availability when compared to antibiotics or inorganic chemicals, they are less toxic and are free of unwanted residues and also act as growth promoters when used as supplements in animal diets (Falcao *et al.*, 2007).

According to Hyun *et al.* (2018) phytochemicals can be used in solid, dried or ground form or as extracts (crude or concentrated) and can also be classified as essential oils and oleoresins depending on the process used to derive the active ingredients (bioactive chemicals). Essential oils especially those that contain phenol and aldehydes, for example cinnamaldehyde, citral, thymol and eugenol as major components could show considerable antibacterial activity (Dormans & Deans, 2000) as presented in Table 1.

Table 1. Common plant extracts, their utilised parts, main active substances and reported properties (Richard, 1992; Charalambous, 1994)

Plant Name	Utilized parts	Main Compounds	Reported properties
<i>Aromatic spices</i>			
Clove	Flower	Eugenol	Appetite Enhancer, Digestive Stimulant, Antiseptic
Cinnamon	Leaf	Cinnamaldehyde	Appetite Enhancer, Digestive Stimulant, Antiseptic
Coriander	Leaf-seed	Linalol	Appetite Enhancer, Digestive Stimulant
Cumin	Seed	Cuminaldehyde	Digestive, carminative, galactagogue
Anise	Fruit	Anethol	Digestion stimulant, galactagogue
Celery	Fruit, Leaf	Phthalides	Appetite and Digestion stimulant
<i>Pungent spices</i>			
Capsicum	Fruit	Capsaicin	Antidiarrhoeic, anti-inflammatory, stimulant, tonic
Pepper	Fruit	Piperine	Digestion stimulant
Ginger	Rhizome	Zingerone	Gastric stimulant
<i>Aromatic herbs and spices</i>			
Garlic	Bulb	Allicin	Digestion stimulant, antiseptic
Rosemary	Leaf	Cineol	Digestion stimulant, antiseptic, antioxidant
Thyme	Whole Plant	Thymol	Digestion stimulant, antiseptic, antioxidant
Sage	Leaf	Cineol	Digestion stimulant, carminative, antiseptic
Peppermint	Leaf	Menthol	Appetite and digestion stimulant, antiseptic

Source: Richard, 1992; Charalambous, 1994.

Plant extracts consists mainly of proteins, peptides, oligosaccharides, fatty acids, vitamins, micro minerals. Plant extracts have a wide range of activities and their active secondary plant metabolites typically belong to the classes of isoprene derivatives and flavonoids (Tajodini *et al.*, 2015). They have a wide range of activities. A great number of plant extracts contain chemical compounds exhibiting antioxidant (Kähkönen *et al.*, 1999; Hashemi *et al.*, 2009), antimicrobial (Hammer *et al.*, 1999; Hsieh *et al.*, 2001),

anti-inflammatory (Pradeep & Kuttan, 2004), anticoccidial (Arab *et al.*, 2006) and anthelmintic (Hoste *et al.*, 2006) properties. The cultivation area, climatic conditions, vegetation phase, genetic modifications and others are factors affecting their chemical and biological diversity (Miliauscas *et al.*, 2004). These properties of plant extract are mainly due to the bioactive compounds such as flavonoids and glucosinolates isoprene derivatives found in nature (Kutlu & Erdogan, 2010). Additionally, the properties probably are the major mechanisms by which plant exert positive effects on the growth performance and health of animals (Hashemi & Davoodi, 2011). They can exhibit their effects by stimulating feed intake and endogenous secretions or having antioxidant, antimicrobial activities as presented in Table 2. All the results obtained depends on the variety of the test materials used, geographical location or environmental condition, species of animals as well as management system.

2. Plant extracts as feed additives in poultry nutrition

Various plant or herbal extracts are commonly included in poultry diets for promoting growth performance and animal health especially when there are health challenging conditions. A lot of researches have documented the beneficial effects of plant extracts on the performance of poultry (Jamroz & Kamel, 2002; Tucker, 2002; Alçıçek *et al.*, 2003; Denli *et al.*, 2004). They reported that the supplementation of plant extracts or oils in to diets increased the body weight gain, feed intake and improved feed conversion rate in poultry (Table 2).

Table 2. Effects of dietary plant extracts used as feed additives on poultry performance

Plant Extract	Dose	Performance effect	Literature
<i>Albizia lebbeck</i> seed oil	0.1-0.4 %	Better growth performance from 7-12 weeks	Alagbe (2019)
Cinnamon	0.2%	Higher growth performance	Al-Kassie, (2009)
<i>Delonix regia</i> extract	5-15 ml/lit	Higher growth in broiler chicks	Alagbe (2019)
Red pepper extract	0.1%	No effect on live performance or in organ morphometrics	Barreto <i>et al.</i> , (2008)
Thyme extracts	3 - 6%	No improved the performance and carcass traits	Amouzmeir <i>et al.</i> , (2002)
Parkia leaf extract	5-20 ml/lit	Increased BW, improved feed efficiency	Alagbe (2019)
Thyme essential Oil	0.2%	Improved growth performance	Danli <i>et al.</i> , (2004)
Cumin seeds	1%	Increased BW, improved feed efficiency	Khalaji <i>et al.</i> , (2011)
Lemon grass + Garlic extract	3-12ml/liter	Higher body weight & feed efficiency	Alagbe & Oluwafemi (2019)
<i>Polyalthia longifolia</i> seed oil	0.2 – 0.4 %	Improved growth performance	Alagbe <i>et al.</i> (2017)

BW: body weight

The main action of plant extracts or oils as feed additives is improving the ecosystem of gastrointestinal microbiota through controlling potential pathogens and digestive capacity in the small intestine (Hashemi & Davoodi, 2011). Wenk *et al.*, (2000) observed that dietary plant extracts exhibit strong effects on stimulate endocrine system and intermediate nutrient metabolism. Tucker (2002) demonstrated a significant

improvement of performance, survive rate in broilers fed diet with many kinds of plant extracts.

Beneficial actions of herbal extracts or their active compounds in animal nutrition may include the stimulation of appetite and feed intake, the improvement of endogenous digestive enzyme secretion (Rahimi *et al.*, 2011). Tollba *et al.*, (2007) reported that at two, four and six week of age, the broiler receiving varying levels of black pepper showed better body weight gain.

Results from the most of studies exhibits beneficial actions of plant extracts on poultry productivity and health. However, some researchers reported that some plant extracts additives have no any effects on growth performance or health status of poultry. Al-Kassie *et al.*, (2011) showed that no difference in broilers fed diet with black pepper. In addition, Aydin *et al.*, (2008) reported that dietary black cumin seed at the level of 1, 2 or 3 % had no effects on body weight gain, feed conversion rate or feed intake in laying hens. Similarly, Hernandez *et al.*, (2004) showed that supplementing broilers diets with essential oil extract from oregano, cinnamon and pepper affected the performance slightly, but this effect was not significant.

3. Conclusions

The use of plant extracts as feed additives in poultry can be valuable because they allow maximizing the overall performance and improvement in digestibility of poultry. The most of plant extracts tested in poultry experiments exhibited positive effects on the productivity and no any harmful effects on animal health and products obtained from the animal. Therefore, they can be used as a botanical alternative in poultry production.

References

- Abd-Elhakeem, A., Abdel -Moty, K., El-Feel, F., & Abu-Elawla, M. (1998). Reproductive performance of buffalo and Friesian calves as affected by flavomycin growth promoter. *Journal of Animal Science*, 35(1), 55-66.
- Alagbe, J.O. (2019). Growth performance and haemato-biochemical parameters of broiler chicken fed different levels of *Parkia biglobosa* leaf extracts. *Academic Journal of Life Sciences*, 5(12), 107-115.
- Alagbe, J.O. (2019). Growth response and bacteria count of broiler starter given *Delonix regia* leaf extract as a natural alternative to antibiotics. *Sumerianz Journal of Agriculture and Veterinary*, 2(9), 76-81.
- Alagbe, J.O., Eimoga, A.A & Alagbe, O.O. (2017). Growth response and carcass characteristics of weaner grasscutters fed diets supplemented with *Polyalthia longifolia* seed oil as a natural growth promoter. *Greener Journal of Agricultural Sciences*, 7(5), 112-119.
- Alagbe, J.O., Grace Funmilayo R. (2019). Effect of *Albizia lebbeck* seed oil dietary supplementation on the haematological and serum biochemical parameters of weaner rabbits. *Sumerianz Journal of Agriculture and Veterinary*, 2(9), 96-100.
- Alagbe, J.O., Oluwafemi, R.A. (2019). Performance and haematological parameters of broiler chicks given different levels of dried lemon grass (*Cymbopogon citratus*) and garlic (*Allium sativum*). *Research in: Agriculture and Veterinary Sciences*, 3(2), 102-111.
- Alçıçek, A., Bozkurt, M. & Çabuk, M. (2003). The effect of a mixture of herbal essential oils, an organic acid or a probiotic on broiler performance. *South African Journal of Animal Science*, 34(4), 217-222.
- Al-Dobaib, S.N. Mousa, H.M. (2009). Benefits and risks of growth promoters in animal production. *Journal of Food, Agriculture & Environment*, 7(2), 202-208.

- Al-Kassie, G.A. (2009). Influence of two plant extracts derived from thyme and cinnamon on broiler performance. *Pakistan Veterinary Journal*, 29(4), 169-173.
- Al-Kassie, G.A., Mamdooh, A.M.A. and Saba, J.A. (2011). The effects of using hot red pepper as a diet supplement on some performance traits in broiler. *Pakistan Journal of Nutrition*, 2(10), 842-845.
- Allam, M., El-Shazly, K., Borhami, B. & Mohamed, M. (2001). Effect of baker's yeast (*Saccharomyces cerevisiae*) supplementation on digestion in sheep and milk response in dairy cows. *Journal of Nutrition and Feeds*, 4, 315-323.
- Amouzmehr, A., Dastar, B., Nejad, J.G., Sung, K.I., Lohakare, J. & Forghani, F. (2012). Effects of garlic and thyme extracts on growth performance and carcass characteristics of broiler chicks. *Journal of Animal Science and Technology*, 54(3), 185-190.
- Angelakis, E., Merhej, V. and Raoult, D. (2013). Related actions of probiotics and antibiotics on gut microbiota and weight modification. *Lancet Infection Diseases*, 13, 889-899.
- Ansan, J.S., Haq, A.U., Yousaf, M., Ahmad, T. and Khan, S. (2008). Evaluation of Different Medicinal Plants as Growth Promoters for Broiler Chicks. *Sarhad Journal of Agriculture*, 24(2), 324-330.
- Arab, H.A., Rahbari, S., Rassouli, A., Moslemi, M.H. and Khosravirad, F. (2006). Determination of artemisinin in *Artemisia sieberi* and anticoccidial effects of the plant extract in broiler chickens. *Tropical Animal Health and Production*, 38(6), 497-503.
- Aydin R., Karaman, M., Cicek, T. and Yardibi, H. (2008). Black cumin (*Nigella sativa* L.) Supplementation into the diet of the laying hen positively influences egg yield parameters, shell quality, and decreases egg cholesterol. *Poultry Science*. 87, 2590–2595.
- Babu, M., Gajendran, K., Sheriff, F.R. and Srinivasan. G. (1992). Crown Grow fit supplementation in broilers improved their performance. *Indian Journal of Poultry Science*, 23, 27-28.
- Barreto, M.S.R., Menten, J.F.M., Racanicci, A.M.C., Pereira, P.W.Z. and Rizzo, P.V. (2008). Plant extracts used as growth promoters in broilers. *Revista Brasileira de Ciência Avícola*. 10(2), 109-115.
- Charalambous, G. (1994). *Spices, herbs and edible fungi*. Elsevier Science Publishers, BV.
- Chen, G., Russell, B. (1999). More monensin-sensitive, ammonia producing bacteria from the rumen. *Applied Environmental Microbiology*, 55, 1052-1055.
- Corpet, D. (1998). The effect of bambermycin, carbadox, chlortetracycline and olaquindox on antibiotic resistance in intestinal coliforms. *Clinical Infectious Diseases*, 135(1), 329-339.
- Deepak, G., Jogi, S., Kumar, A. Bais, R., & Vikas, K.S. (2002). Effect of herbal liver stimulants on efficacy of feed utilization in commercial broiler chicken. *Indian Journal of Animal Research*, 36(1), 43-45.
- Denli, M., Okan, F. & Uluocak, A.N. (2004). Effect of dietary supplementation of herb essential oils on the growth performance, carcass and intestinal characteristics of quail (*Coturnixcoturnix japonica*). *South African Journal of Animal Science*, 34, 174-179.
- Fulton, R.M., Nersessian, B.N. & Reed, W.M. (2002). Prevention of *Salmonella enteritidis* infection in commercial ducklings by oral chicken egg derived antibody alone or in combination with probiotics. *Poultry of Science*, 81, 34-40.
- Gaines, A., Rollins, L. & Selwyn, M. (1990). Effect of penicillin and virginiamycin on drug resistance in lactose-fermenting enteric flora. *Anti-microbial Agents and Chemotherapy*, 17, 428-433.
- Ganguly, S. (2013). Herbal and plant derived natural products as growth promoting nutritional supplements for poultry birds: A Review. *Journal of Pharmaceutical and Scientific Innovation*, 2(3), 12-13.
- Ganguly, S., Prasad, A. (2010). Role of plant extracts and cow urine distillate as immunomodulator in comparison to levamisole. *Journal of Immunological Immunopathology*, 12(2), 91-94

- Gunther, K.D. (1994). The role of probiotics as feed additives in animal nutrition. In Piva, G. (ed.). Proc. IV Inf. Feed Production Conference, Piacenza, pp. 97-114
- Guo, F.C., Williams, B.A., Kwakkel, R.P., Li, H.S., Li, X.P., Luo, J.Y., Li, W.K. & Verstegen, M.W.A. (2004). Effects of mushroom and herb polysaccharides, as alternatives for an antibiotic, on the cecal microbial ecosystem in broiler chickens. *Poultry Science*, 83(2), 175-182.
- Hammer, K.A., Carson, C.F. & Riley, T.V. (1999). Antimicrobial activity of essential oils and other plant extracts. *Journal of Applied Microbiology*, 86(6), 985-990.
- Hashemi, S.R., Davoodi, H. (2010). Herbal plants and their derivatives as growth and health promoters in animal nutrition. *Veterinary Research Communications*, 35(3), 169-180.
- Hernandez, F., Madrid, J., Garcia, V., Orengo, J. & Megias, M.D. (2004). Influence of two plant extracts on broilers performance, digestibility, and digestive organ size. *Poultry Science*, 83(2), 169-174.
- Hoste, H., Jackson, F., Athanasiadou, S., Thamsborg, S.M. & Hoskin, S.O. (2006). The effects of tannin-rich plants on parasitic nematodes in ruminants. *Trends in Parasitology*, 22(6), 253-261.
- Hsieh, P.C., Mau, J.L. & Huang, S.H. (2001). Antimicrobial effect of various combinations of plant extracts. *Food Microbiology*, 18(1), 35-43.
- Jamroz, D., Kamel, C. (2002). Plant extracts enhance broiler performance. In Non-ruminant nutrition: Antimicrobial agents and plant extracts on immunity, health and performance.
- Jamroz, D., Wiliczkiewicz, A., Wertelecki, T., Orda, J. and Skorupińska, J. (2005). Use of active substances of plant origin in chicken diets based on maize and locally grown cereals. *British Poultry Science*, 46(4), 485-493.
- Kähkönen, M.P., Hopia, A.I., Vuorela, H.J., Rauha, J.P., Pihlaja, K., Kujala, T.S. and Heinonen, M. (1999). Antioxidant activity of plant extracts containing phenolic compounds. *Journal of Agricultural and Food Chemistry*, 47(10), 3954-3962.
- Khalaji, S., Zaghari, M., Hatami, K.H., Hedari-Dastjerdi, S., Lotfi, L. and Nazarian, H. (2011). Black cumin seeds, Artemisia leaves (*Artemisia sieberi*), and *Camellia* L. plant extract as phytogenic products in broiler diets and their effects on performance, blood constituents, immunity, and cecal microbial population. *Poultry Science*, 90(11), 2500-2510.
- Krishan G., Narang A. (2014). Use of essential oils in poultry nutrition: A new approach. *Journal of Advanced Veterinary and Animal Research*, 1(4), 156-162.
- Kumar, O.M. (1991). Effect of Liv-52 syrup on broiler performance in North Eastern region, *Indian Journal of Poultry Science*, 37-38.
- Kutlu, T., Erdoğan, Z. (2010). Kanatlı Beslemede Fitobiyotik Yem Katkı Maddeleri. Kümes Hayvanları Kongresi, 07-09 Ekim, Kayseri.
- Lillehoj, H., Liu, Y., Calsamiglia, S., Fernandez Miyakawa M., Chi F., Cravens R., Oh S. & Gay C. (2018). Phytochemicals as antibiotic alternatives to promote growth and enhance host health. *Journal of Veterinary Research*, 10(18), 49-76.
- Manzanilla, E.G., Baucells, F., Kamel, C., Morales, J., Perez, J.F. and Gasa, J. (2001). Effects of plant extracts on the performance and lower gut microflora of early weaned piglets. *Journal of Animal Science Supplement*, 1, 473-474.
- Mee, B. (1992). The selective capacity of pig feed additives and growth promotants for coliform resistance. *Journal of Antibodies and Agriculture*, 4, 349-358.
- Miliauskas, G., Venskutonis, P.R. & Van Beek, T.A. (2004). Screening of radical scavenging activity of some medicinal and aromatic plant extracts. *Food Chemistry*, 85(2), 231-237.
- Mishra, S.J., Singh. D.S. (2000). Effect of feeding root powder of *Withania somnifera* (L.) Dunal (aswagandha) on growth, feed consumption, efficiency of feed conversion and mortality rate in broiler chicks. *Bioved. (annual)*, 11, 79-83.

- Pradeep, C.R. and Kuttan, G. (2004). Piperine is a potent inhibitor of nuclear factor- κ B (NF- κ B), c-Fos, CREB, ATF-2 and pro-inflammatory cytokine gene expression in B16F-10 melanoma cells. *International Immunopharmacology*, 4(14), 1795-1803.
- Richard, H.J., Nolean, I. and Giampoli, P. (1992). Techniques of analysis of the spices and aromatics. Epicesetaromates (Richard H., ed). Tec & Lavoisier, Paris. 191-211.
- Tajodini, M., Saeedi, H.R. and Moghbeli, P. (2015). Use of black pepper, cinnamon and turmeric as feed additives in the poultry industry. *World's Poultry Science Journal*, 71(1), 175-183.
- Tollba, A.A.H., Wagdy, A.Z. and Shabaan, S.A.M. (2007). Improvement of fayoumi laying hens performance under hot climate conditions. 1-probiotic and prebiotic. Egypt. *Poultry Science*, 27, 1-20.
- Tucker, L. (2002). Botanical broilers: Plant extracts enhance broiler performance. *Feed International*, 23(9), 26-29.
- Wang, R., Li, D. & Bourne, S. (1998). Can 2000 years of herbal medicine history help us solve problems in the year 2000? Biotechnology in the feed industry: Proceedings of Alltech's 14th Annual Symposium, Kentucky, USA. 273-291.
- Wenk, C. (2000). Why all the discussion about herbs? Biotechnology in the feed industry. Proc. Of Alltech's 16 thAnnu. Symp. Alltech Technical Publications, Nottingham University Pres. Nicholasville, KY, 79-96.